



# **AN INVESTIGATION INTO THE CORRELATION BETWEEN SPECIFIC GENETIC MUTATIONS AND COLISTIN RESISTANCE IN *ESCHERICHIA COLI***

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# **BACKGROUND OF COLISTIN**

- Last line of defence for treating multi-drug resistant Gram negative bacterial infections
- Cationic polypeptide with a fatty acid chain attached
- Targets the lipopolysaccharide molecule
- Use in human medicine becoming more common

# COLISTIN IN MEDICINE

- Manufactured as colistin sulfate or colistimethate sodium
- Colistin sulfate used in bowel decontamination and treating surface infections
- Colistimethate sodium used to treat MDR *Pseudomonas aeruginosa* infections in cystic fibrosis patients
- Associated with nephrotoxicity and neurotoxicity

# CHROMOSOMAL COLISTIN RESISTANCE

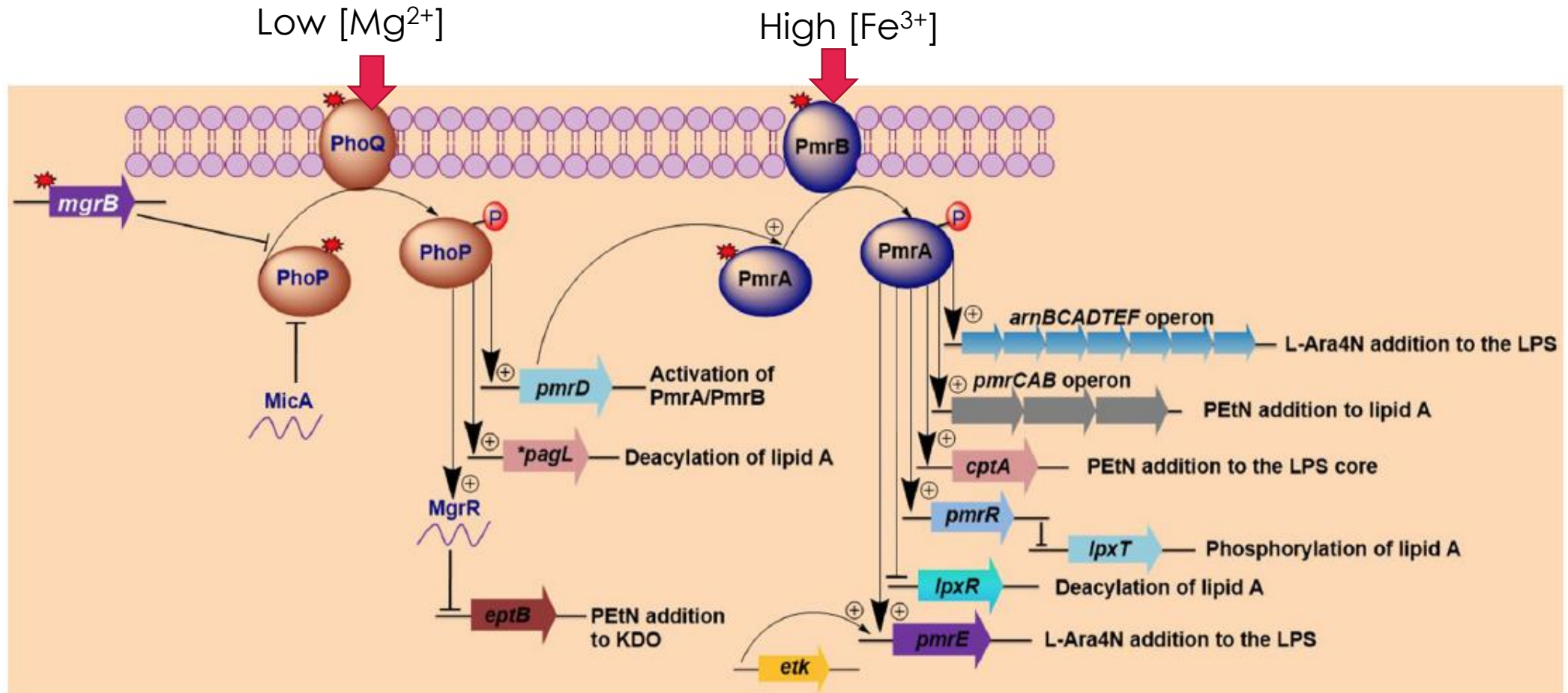


Figure 1: Colistin resistance mechanisms (1)

# PROJECT METHODOLOGIES

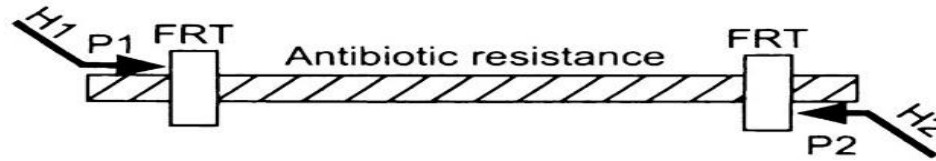
1. Microbroth Dilution to investigate antibiotic minimum inhibitory concentrations

2. Lambda Red Recombination for deleting *phoP* from *E. coli* ATCC 25922 genome

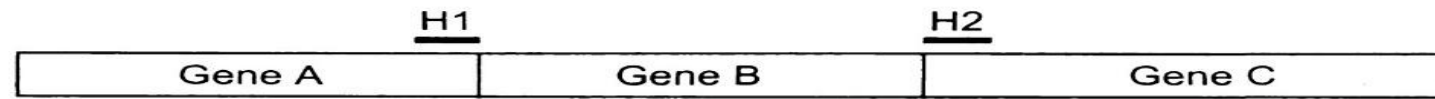


# 2. LAMBDA RED RECOMBINATION

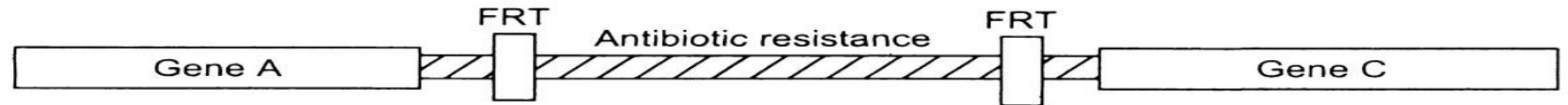
Step 1. PCR amplify FRT-flanked resistance gene



Step 2. Transform strain expressing  $\lambda$  Red recombinase



Step 3. Select antibiotic-resistant transformants



Step 4. Eliminate resistance cassette using a FLP expression plasmid

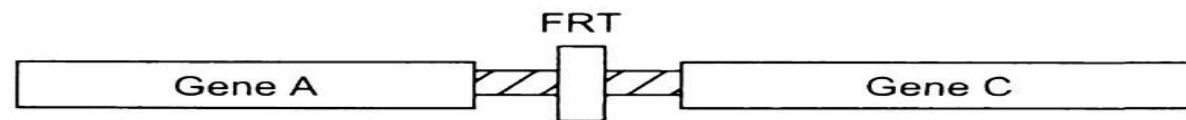


Figure 2: Schematic demonstrating Lambda Red Recombination (2)

**Table 2: *mcr-1* status and colistin MIC interpretation of *E. coli* test isolates and control strain**

<b>Isolate</b>	<b><i>mcr-1</i> Status of Isolate</b>	<b>Colistin MIC Interpretation</b>
<b>M6</b>	<b><i>mcr-1</i> negative</b>	<b>RESISTANT</b>
<b>M73</b>	<b><i>mcr-1</i> negative</b>	<b>RESISTANT</b>
<b>M33</b>	<b><i>mcr-1</i> positive</b>	<b>LOW LEVEL RESISTANCE</b>
<b>M52</b>	<b><i>mcr-1</i> positive</b>	<b>SENSITIVE</b>
<b>M4</b>	<b><i>mcr-1</i> negative</b>	<b>SENSITIVE</b>
<b>M44</b>	<b><i>mcr-1</i> negative</b>	<b>SENSITIVE</b>
<b>Control ATCC 25922</b>	<b><i>mcr-1</i> negative</b>	<b>SENSITIVE</b>



# VERIFICATION OF *phoP* DELETION

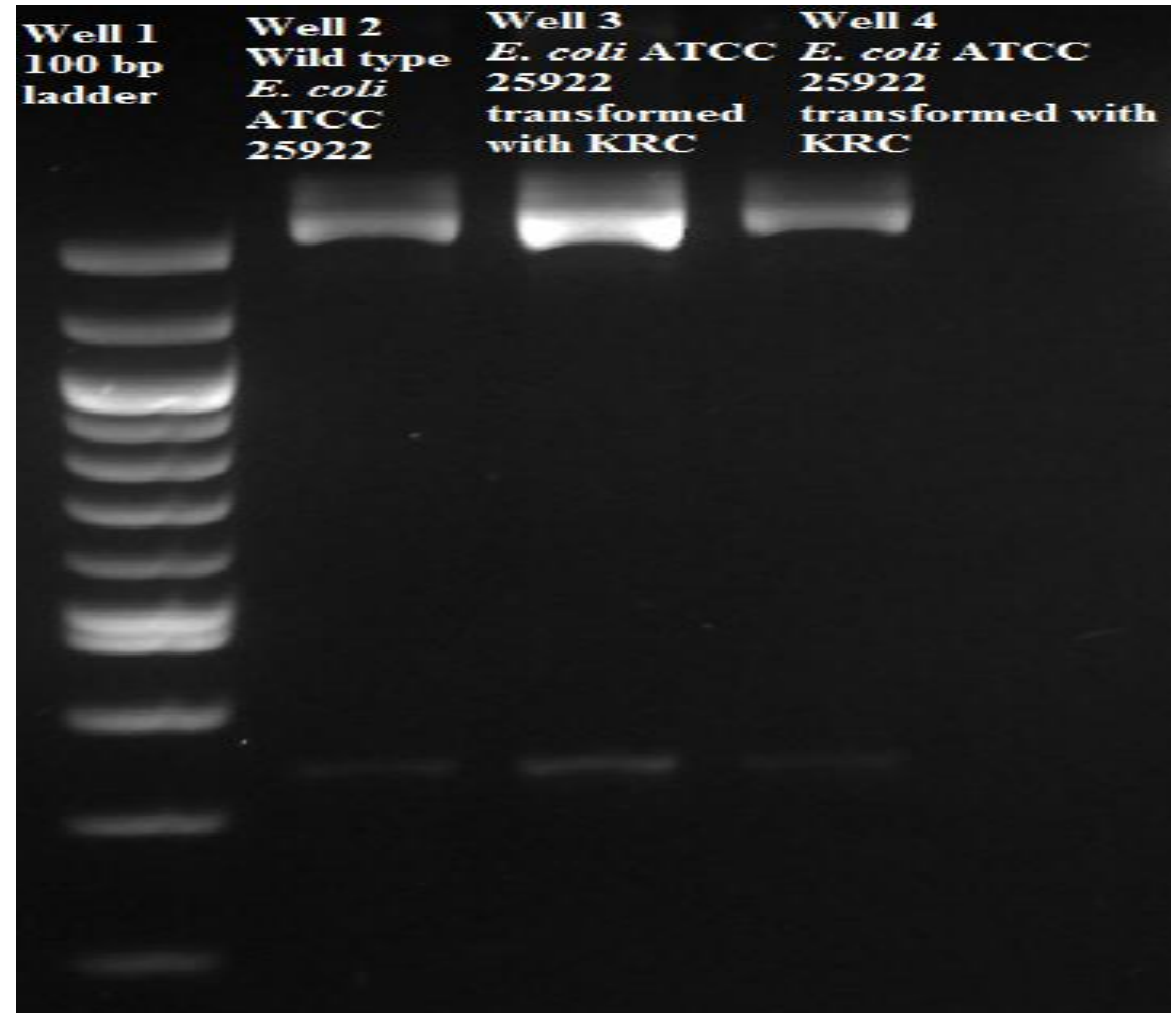


Figure 4: Electrophoresis demonstrating the presence of kanamycin resistance cassette in Well 2 and *phoP* in Wells 2 and 4

# VERIFICATION OF KRC DELETION



*Figure 5*: Electrophoresis graph demonstrating the successful deletion of the KRC from mutant *E. coli* ATCC 25922

**Table 3: Colistin MIC of wild type *E. coli* ATCC control strain and mutant control strain**

	<u>Colistin MIC (mg/l)</u>	<u>Interpretation</u>
<b><i>Wild type E. coli</i> <i>ATCC 25922</i></b>	0.125	<b>SENSITIVE</b>
<b><i>Mutant E. coli</i> <i>ATCC</i> <i>25922</i></b>	<u><b>0.25</b></u>	<b>SENSITIVE</b>

# CONCLUSION

- One fold increase in MIC value is significant
- Demonstrates other two-component systems compensate for loss of *phoP* upon colistin exposure
- Also implies *phoP* mutation alone cannot mediate colistin resistance
- Multiple mutations in two-component systems are required for the development of a colistin resistant phenotype

## **FUTURE WORK**

- Clone *mcr-1* into:
  - Susceptible *E. coli* control strain ATCC 25922
  - Mutant *E. coli* ATCC 25922
- Investigate the true function of the *mcr-1* gene – does it actually confer colistin resistance or serve to augment colistin resistance mechanisms?

## **REFERENCES**

- (1) Olaitan AO, Morand S, Rolain J-M. Mechanisms of polymyxin resistance: acquired and intrinsic resistance in bacteria. *Frontiers in Microbiology*. 2014; 5: 634.
- (2) Datsenko KA, Wanner BL. One-step inactivation of chromosomal genes in *Escherichia coli* K-12 using PCR products. *Proceedings of the National Academy of Sciences of the United States of America*. 2000; 97(12): 6640 – 6645.

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